conducted 40 total hours of observations of 8 EVS personnel before and 10 after the product substitution. Mean compliance scores increased from 14.3 to 18.2 after the substitution. Noncompliance when dilutable products were used was most often due to incorrect use of products, including inappropriate mixing of disinfectant products (e.g., dilutable quaternary ammonium plus sodium hypochlorite) and inadequate contact time. During post-discharge and daily cleaning before the substitution, activities other than cleaning and disinfection accounted for 50% and 55% of total EVS personnel time, respectively (Figure), with similar time distribution after the substitution. **Conclusions:** Switching from dilutable to RTU disinfectants improved compliance with recommended cleaning and disinfection practices. EVS personnel spend a substantial proportion of their time doing activities other than cleaning and disinfection.

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Presentation Type:
Poster Presentation
Subject Category: Disinfection/Sterilization
Evaluation of a continuously active surface disinfectant against Candidozyma auris‡
Rachel Medernach[1], Mackenzie May, Ellen Gough[1], Sarah Sansom[2] and Mary Hayden[1]
[1]Rush University Medical Center and [2]Rush University

Background: Candidozyma auris is an emerging healthcare-associated pathogen that colonizes human skin and survives on healthcare surfaces for up to two weeks. As C. auris re-contaminates the healthcare environment within hours of disinfection, a continuously active disinfectant may provide an advantage over traditional disinfectants in healthcare environments with high rates of C. auris colonization. Water-stable organosilanes (WSOs) have demonstrated continuously active disinfection against bacterial and viral species but have not been studied against C. auris. Goldshield® GS75 (Locust Valley, NY) is a WSO composed of a siloxane bonding agent providing surface coating, a nitrogen molecule for organism attraction, a long carbon chain for cellular penetrance, and a quaternary ammonium compound for disinfection. We tested the efficacy and duration of effect of GS75 on survival of C. auris in vitro. Methods GS75 was applied to C. auris colony forming units (CFUs). After one hour, coupons

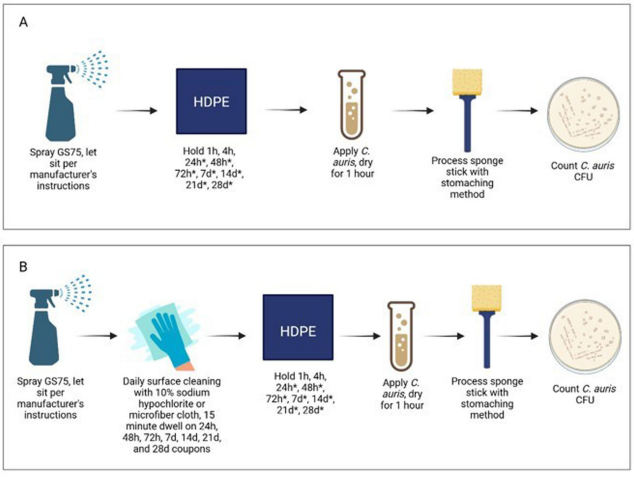


Figure: Experimental design. A) Testing efficacy and duration of effect of GS75 on *C. auris* survival; B) Testing efficacy and duration of effect of GS75 on *C. auris* survival, with addition of wiping with a dry microfiber cloth or bleach wipe. While use of a continuously active disinfectant should reduce the need for routine cleaning and disinfection in a healthcare setting, we anticipate that some additional cleaning and disinfection will be needed. Experiments done in triplicate. *=24-hour re-inoculation with *C. auris* to simulate daily shedding, with sponge stick collection occurring on denoted days one hour after inoculation. HDPE, high-density polyethylene; CFU, colony forming units.

Length of time	Average CFU ± SD			
	Untreated	GS75 treated only	GS75 treated + daily bleach	GS75 treated + daily microfiber
1 hour	283 ± 35	0.44 ± 0.73	4 ± 3	9 ± 3
4 hours	413 ± 38	5 ± 2	0.56 ± 1	14 ± 3
24 hours*	146 ± 8	0.11 ± 0.33	0.44 ± 0.73	0.67 ± 0.50
48 hours*	577 ± 93	9 ± 6	3 ± 2	3 ± 2
72 hours*	257 ± 81	19 ± 6	1 ± 2	8 ± 2
7 days*	410 ± 78	1 ± 1	3 ± 2	1 ± 2
14 days*	590 ± 61	5 ± 3	4 ± 6	12 ± 3
21 days*	300 ± 20	4 ± 2	6 ± 7	26 ± 6
28 days*	350 ± 36	6 ± 4	0 ± 0	13 ± 6

Table: Mean colony forming units (CFUs) detected at each time point, with all GS75 conditions noting reduced *C. auris* growth compared to the untreated condition. SD=standard deviation. *=denotes time periods where *C. auris* was inoculated daily to mimic repeated contamination

were swabbed using premoistened sponge-sticks (Neogen Sponge-Stick with proprietary neutralizing buffer active against chlorine and QACs; Neogen, Lansing, MI), processed using the stomacher method and plated quantitatively to Sabouraud Dextrose agar (RemelTM, Lenexa, KS) then incubated for 48 hours at 37°C in ambient air. Duration of GS75 activity was assessed using the same method, with C. auris re-inoculated daily to simulate repeated contamination from a C. auris colonized or infected patient, and recovery attempted at 4 hours, 24 hours, 48 hours, 72 hours, 7 days, 14 days, 21 days, and 28 days after coupon treatment. (Figure). A separate set of coupons was wiped daily with a dry microfiber cloth or a 10% sodium hypochlorite bleach wipe (Sani-Cloth®, PDI, Woodcliff Lake, NJ) before daily re-inoculation to simulate routine environmental cleaning practices that might be necessary even with use of a continuously active disinfectant. Results Mean C. auris CFUs recovered from untreated coupons exceeded 145 at all sampling times, while GS75-treated coupons showed significantly reduced C. auris recovery. The addition of daily bleach or microfiber cloth wiping had minimal effect on GS75 activity (Table). Conclusion One application of GS75 reduced survival of C. auris on a common healthcare facility surface material for up to 28 days, despite repeated C. auris challenge and with minimal reduction in activity after physical or chemical cleaning. These findings support evaluation

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Poster Presentation
Subject Category: Emerging Pathogens
Antibiotic Susceptibility Patterns of Extended-Spectrum Beta-Lactamase-Producing Enterobacterales (ESBL-E), Tennessee 2019–2024‡
Sarah Petersen[1], Daniel Muleta[1], Christopher Evans[2], Cherlly Bailey[3], Carissa Wallace[4] and Becky Meyer[1]
[1]Tennessee Department of Health; [2]TN Department of Health; [3]TN Dept of Health and [4]TENNEESE DEPARTMENT OF HEALTH

Background: Extended-spectrum beta-lactamase-producing Enterobacterales (ESBL-E) are a growing public health concern. This study investigates antibiotic resistance patterns for ESBL-E (Escherichia coli and Klebsiella species) cases in Maury and surrounding counties in Tennessee as part of the Emerging Infection Program's Multi-site Gram-negative Surveillance Initiative. **Methods:** ESBL-E cases are defined as extended-spectrum cephalosporin-resistant E. coli, Klebsiella pneumoniae, Klebsiella variicola, or Klebsiella oxytoca isolated from urine or normally sterile sites from residents of the surveillance area. Cases from July 2019 through December 2024 were included. Susceptibility data were collected from clinical laboratory reports. E. coli isolate resistance was compared to aggregated Klebsiella species isolate resistance using χ or fisher's exact tests. Organism specific yearly resistance trends excluded the incomplete 2019 collection year and were examined via simple linear regression.

‡The abstract titles in this supplement have been corrected since original publication. A notice detailing the changes has been published at https://doi.org/10.1017/ash.2026.10802.

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