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Artificial Intelligence Prospective

Automating material image analysis for material discovery

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Abstract

Advancements in temporal and spatial resolutions of microscopes promise to expand the frontiers of understanding in materials science. Imaging techniques produce images at a high-frame rate, streaming out a tremendous amount of data. Analysis of all these images is time-consuming and labor intensive, creating a bottleneck in material discovery that needs to be overcome. This paper summarizes recent progresses in machine learning and data science for expediting and automating material image analysis. The discussion covers both static image and dynamic image analyses, followed by remarks concerning ongoing efforts and future needs in automated image analysis that accelerates material discovery.

Introduction

Material scientists have long desired to image, with high-spatial and temporal resolutions, material structures, and interactions in chemical, material, and biologic processes, because visual information with sufficient details is vital to discovery as emphasized in many scientific activities.[1] Major research efforts have been made to develop material imaging tools, including various kinds of electron microscopes (EM)[2] and scanning probe microscopes (SPM).[3] The wide use of these microscopes leads naturally to the exponential increase in material imagery data, which in turn calls for efficient and effective analysis methods to extract useful information from the raw images for aiding designs, discoveries, and decision-makings in materials science.

An EM operates with an electron beam, which illuminates materials to create an image of materials. A scanning electron microscope (SEM) creates images based on electrons reflected by material surfaces, whereas a transmission electron microscope (TEM) creates images based on electrons transmitted through material samples. When the electron beam in a TEM rasters across the sample, the resulting technique is called scanning transmission electron microscopy (STEM). Historically, EM techniques were capable of capturing only a static image, because an EM requires samples to be placed in a high-vacuum environment, meaning that the samples are typically frozen or dried before imaging. Recent progress in environmental *in situ* EM, including liquid phase EM and *in situ* gas EM, allows direct, high magnification, and real-time imaging of material structures or objects during experiments, capturing their evolution at high-imaging speeds.[4–11] By their very nature, these imaging strategies generate material images at a high-frame rate, streaming out a tremendous amount of image data.

An SPM makes use of the dynamic interaction between a cantilever and a material, for inferring the nanoscale surface structure and functionality of the materials. Considerable progress in SPM techniques has been made since early 2000s. In particular, there is a growing trend toward developing multi-frequency SPM techniques that enable simultaneous capture of multiple imaging channels and complex modes of operation.[12,13] These new techniques generate a large volume of multi-dimensional data, creating needs for novel image analyses that visualize and convert the raw data to material specific information. Machine learning research for SPM data is reviewed in Kalinin's recent review paper.[14] This review paper focuses therefore on machine learning and data science research for EM data.

Research needs for machine learning and data science in high-resolution imaging of materials are the topic of increasing discussion in many nation-wide research initiatives and white papers, including the National Nanotechnology Initiative's Signature Initiative[15] and the recent Department of Energy (DOE) workshop report,[1] entitled "Basic Research Needs For Innovation and Discovery of Transformative Experimental Tools," emphasizing new machine learning and data science methods to triage, accelerate, and scale up the analysis of image data to provide real-time feedback to materials science researchers. Material image data are of large volumes because of the high-frame rates and high-spatial resolutions required to capture interesting phenomena, as explained above. There is also an increasing desire to search image data with great attention to find the right kind of details, because materials frequently embed complicated structural features that may not be obvious in a plain view. There are unresolved issues in almost every facet of the material image processing problems, ranging

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from data storage and communication to analysis and decision making.

Manual analysis of high-resolution image data is definitely inefficient and may no longer be feasible in the era of big data. The manual task could take months of dedicated work by an expert just for a partial analysis. The fast arrival of large volumes of material image data creates a major bottleneck in advancing fundamental materials science, in spite of the fact that the innovation in imaging techniques is an enabler in the first place leading to new material discoveries. What this means is that in addition to the development of image acquisition techniques and instruments, we are in pressing needs of better and capable image data analysis methods to convert the massive amounts of raw data into useful information that is directly or substantively linked to fundamental chemical and physical mechanisms of material formation and dynamics.

Motivated by the emerging needs, this paper reviews recent research efforts that aim at automating material image analysis. Our review is organized based on different perspectives of visual information relevant to materials research, including morphology, location, dispersion, and spatial arrangements of materials, and dynamic material evolutions. We also discuss ongoing efforts and future research needs, following the review.

Morphology analysis

Many materials science problems are centered around understanding the relationship between material structures and properties or functionalities. Material structures are often imaged and characterized using microscopy techniques, and are described by the outlines of material interior and exterior structures as well as the geometric features of these outlines, such as sizes, shapes and topology. We call the problem of extracting the outlines from material images and quantifying their geometric features as *morphology analysis*. We discuss the state-of-the-art in morphology analysis for two-dimensional (2d) microscopic images and extend the discussion to the three-dimensional (3d) images in the last paragraph of this section.

The most challenging part in morphology analysis is to extract the outlines of the material structures. The task is difficult due to many complicating factors, including but not limited to, the high image noises, low image contrasts, and uneven background illumination. Many practitioners attempted to apply simple edge detection algorithms such as Canny's edge detector[16] that locates the "edge pixels" or the intensity jump pixels around the interfacial areas between different materials. For micrographs of low contrast, however, "true" edge pixels and the intensity jumps become almost indiscernible due to background noises, and consequently, the edge detector produces fragmented edges, mixed with noise pixels. Combining the fragmented edge pixels into holistic outlines is not straightforward.

When the outlines of the materials imaged do not overlap, a good alternative is to denoise and binarize an image to its background and foreground—foreground corresponding to materials—and then extract the outlines of the foreground, as illustrated in Fig. 1. This approach is referred to as *image binarization*. The simplest approach for image binarization is a global image thresholding that compares image pixel intensities with a chosen threshold; a popular algorithm which most commercial or open source software implements is the Otsu image thresholding.[17] However, the algorithm does not work well for many micrographs, particularly when the background intensity of a micrograph changes in space due to focused beam radiation or transitions between different background materials, to which local thresholds need to be applied.[18] Local thresholding is to estimate the background of an image, subtract the background from the image, and then apply a global threshold on the subtracted image (i.e., the residual image); the steps and intermediate results are illustrated in Figs. 1(b)–1(d). Vo and Park[19] provide a comprehensive summary of image binarization approaches and propose an effective background estimation method, enabling local thresholds to be constructed for a robust binarization of material images.

When materials are overly crowded in a sample and their images in a micrograph partially overlap, a simple binarization of the micrograph gives only the outlines of the overlapped materials instead of individual outlines. More sophisticated image analysis methods are needed to recover the outlines of individual material objects or structures buried in the overlaps. Park et al.[20,21] framed three necessary analysis steps for extracting the individual outlines. The first is to segment the binarization outcome into the images of individual material objects. In computer vision, such separation step is called *image segmentation*.[22] The segmentation by itself does not yield the complete outlines of individual material objects, because the outlines are partially occluded due to the overlapping, and as such, the segmentation outcomes are incomplete outlines as illustrated in Fig. 2(c). The missing parts of the outlines must be inferred and recovered; this step is referred to as *outline inference* and illustrated in Fig. 2(d). The inference can exploit prior knowledge of material structures to fill in the blanks. For example, in particle analysis, when nanoparticle shapes are circular or elliptical, the prior shape information is used to formulate the segmentation and outline inference problem.[23,24] When the prior knowledge is unavailable, material structure information of the well-isolated material structures in the same sample can be exploited for inference. For doing so, statistical modeling is needed to encode the structural information, which includes defining the metric and probability space of shapes or structures and formulating the statistical inference problems on the shape space. Park et al.[21] used a curve, parameterized by multivariate parameters, to represent a shape. A Gaussian mixture distribution was imposed on the parameters to represent multiple different shapes of materials and their variations. The modeling was combined with the outline inference step, which was iteratively solved by an expectation-conditional-maximization algorithm.[25] For very noisy images taken from solid samples, complementary imagery information such as both intensity and gradient must be

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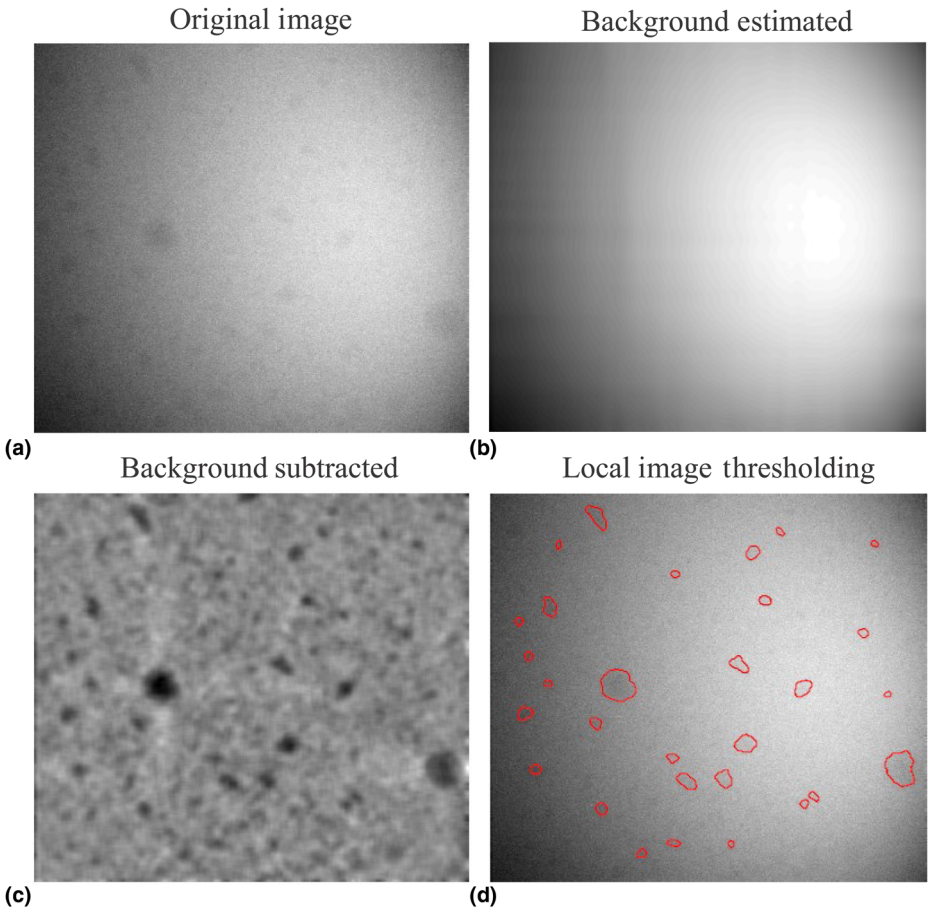


Figure 1. Image binarization to extract the outlines of nanocrystals: (a) original image, (b) background image estimated, (c) image after the background subtraction, and (d) outlines extracted by local image thresholding.

used together to achieve a more practical segmentation and shape identification.[23] Nevertheless, the current approaches tend to use a curve representation of material outlines, which may not apply to the outlines of materials having complex geometries and topology. A general shape space needs to be defined. This calls for new research ideas and approaches in modern shape data analysis and topological data analysis.[26] A Bayesian object identification[27] can presumably handle

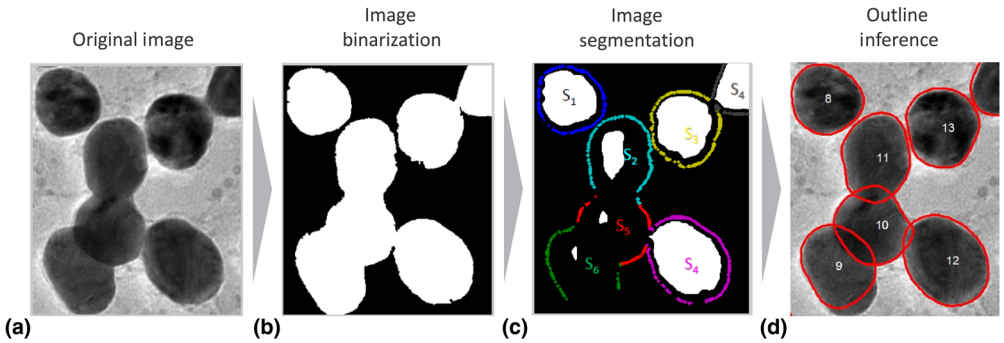


Figure 2. (a) Three steps for morphology analysis; (b) the original image is binarized for the binary image of partially overlapping materials in the image; (c) the binary image is segmented into individual foreground images; and (d) the occlusions due to overlaps are estimated and filled for complete outlines. *Reprinted with permission from Park et al.[21]

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complex shapes from a library of prescribed shapes, but faster Bayesian solution algorithms need further attention and research.

All the aforementioned morphology analysis methods have been mainly devised for extracting and analyzing the outlines of material objects in 2d microscopic images, for the understandable reasons that the majority of material imaging instruments are still 2d microscopy techniques. Although 3d electron tomography does exist and is gaining popularity,[28] electron tomography comes with many more unresolved technical issues; a case in point is the well-known missing wedge problem due to the limited tilt angle series of sinograms.[29] Morphology analysis in 3d would involve similar analysis steps, outlined above for the same analysis in 2d,[21] but each step is much more complicated due to the complexity in the representation of 3d structures. The aforementioned 2d morphology analysis methods are by and large based on a curve representation of material boundary, which is ineffective in representing 3d morphologies. Should the same 2d models be used for 3d objects, a much greater degree of freedom would have to be used, but then, the models would become too complex to be practical. Furthermore, many 3d morphologies come with intrinsic topologies with interior holes, which cannot be represented adequately by commonly used 2d surface or shape models. The lack of effective representation of shapes and topologies in 3d is still a big hurdle to having effective 3d morphology analyses.

Location, dispersion, and lattice analysis

Beyond morphology, studies on the spatial positioning and arrangements of smaller scale elements within bulk scale materials are of great interest to material scientists because analysis of such arrangements could yield insights concerning functionalities of materials. For example, crystallographic studies at atomic length scales locate individual atoms and identify symmetries, dislocations, and defects in their locations. Quantification of these material features allows material scientists to map material performance as a function of measurable parameters such as distance or energy, which can be extracted directly from microscopic data.[30] Another example is the composite matrix mixed with nanoparticles. Properties of a composite, such as its strength, conductivity, or transparency, can be remarkably enhanced by blending nanoparticles into the polymer host materials, but the result of the enhancement is inevitably dependent on the dispersion and distribution of nanoparticles within the host materials.[31] Here we summarize the state-of-the-art machine learning and data science approaches developed in these two specific contexts.

The progress in high-resolution, real space imaging techniques has allowed 10 pm or better precision in the measurements of atomic positions. The amazing technology opens new scientific discovery for correlating the lattice structures of atoms and the lengths and angles of bonds to the functionalities of materials.[32] Typical image analysis for such scientific research comes in two stages, i.e., locating individual atom locations and identifying local and global symmetries, or deviation from symmetries, i.e., defects in locations. The first stage of the analysis without any local or global symmetry references is a general spot detection problem. Although spot detection algorithms exist, such as Laplacian of Gaussian filters,[33–35] employing them to locate atoms is a non-trivial matter because those contrast-based algorithms are prone to errors, particularly for low-contrast images (which is common in material imagery). Once all atom locations are identified, location symmetries or the lack thereof (defects) are characterized by analyzing the spatial arrangement of the atom locations. The case for all atoms being homogeneous and aligned to a global lattice with hardly any defects has been extensively studied, and such structure can be described by a global descriptor, i.e., the basis vectors of the global lattice, as illustrated in Fig. 3. When materials are composing of multiple heterogeneous atoms and multiple local lattice structures, the global descriptor is insufficient to describe the material structure. Consequently, multiple local descriptors are needed to describe such multiphase materials. Currently, there is no well accepted approach to describe such varying, heterogeneous local structures. Some papers analyze the spatial arrangements of the nearest neighboring atoms using distance matrices and the corresponding principal components.[32,36] A principal score is assigned to each atom and visualized for finding the boundaries among different phases. More recent research tries to solve the two-stage problem (identification and characterization) simultaneously, instead of doing so separately and sequentially.[30] Intuitively, the outcome from one stage could help improve the accuracy of analysis at the other stage. For instance, knowing the local and global symmetric patterns of atoms definitely helps improve the accuracy of identifying individual atom locations in the low-contrast images, because the symmetry information can be used as a reference to hint on potential atom locations.

Analyzing the mixing states of nanoparticles in a composite material is of great interest due to the relationship between the mixing states and the resulting material functionalities. The

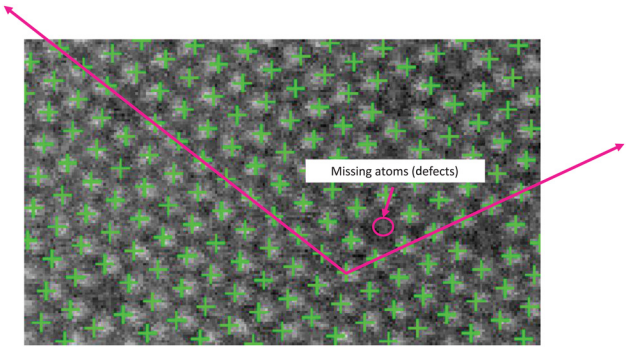


Figure 3. Symmetries and defects of atom locations. The crosses point individual atom locations, and two arrow lines represent the basis vectors of a global lattice. *Figure from Li et al.[30]

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mixing states are described using the metrics known as dispersion and distribution. Dispersion refers to the degree of nanoparticles being separated instead of aggregating, whereas distribution refers to the degree of nanoparticles being located uniformly throughout the host composite material. The characterization of the mixing states of a composite mixture was performed by taking and analyzing the electron microscopic image of the composite mixture. When nanoparticles are well dispersed in a composite, the centroids of individual nanoparticles can be extracted, and the distribution of the centroids is used to quantify the distribution of nanoparticles. The Ripley's K function[37] is a spatial statistic that quantifies the homogeneity of spatial point distribution, which was applied to the centroid locations for characterizing the distribution of nanoparticles.[38] Direct application of the spatial statistics such as the Ripley's K function loses its effectiveness when nanoparticles aggregate severely, forming large agglomerates in a composite. The spatial statistical approaches treat each material object as a dimensionless dot in the space. In doing so, the approach counts an agglomerate of multiple particles as a single particle, losing the information of the individual particles. One can certainly discretize each agglomerate with patches of smaller sizes that are comparable with that of individual particles, and then apply the Ripley's K function to the centroids of patches. A great care[31] must be taken to adjust the normalizing constant used in the Ripley's K function or other similar spatial statistics metrics; otherwise, the resulting analysis could lead to erroneous orders while ranking and selecting the material samples with the best mixing state.

Dynamic material evolution

Many material and biologic processes take place in a liquid or gas environment, where the final process outcome is the result of a complicated transient process in which the material objects evolve from its initial state to the equilibrium state. Understanding and subsequently controlling the final process outcome require a capability of directly observing the kinetics of the transient processes as they are happening. Recent advances in *in situ* and *operando* electron microscopy allow atomic resolution imaging of the transient processes with high temporal resolutions, promising to expand the frontiers of our understanding in material and biologic science.[39–41] One example is the bottom up colloidal self-assembly process in which nanoparticles are organized into ordered structures.[42] *In situ* microscopes take motion pictures of the nanoparticles as they are nucleating, aggregating, and morphing into different sizes and shapes. Another example is in cell biology. Microtubules are the main organizational system in cells, responsible for cell shape, intracellular transport, and cell division.[43] One of the mysteries of cell biology is the basic set of properties involved in the seeding and growth of microtubules and the processes underpinning how the systems are initiated.[44] *In situ* microscopes make it possible for scientists to directly image the underlying dynamic processes for their fundamental science studies.

By their very nature, the *in situ* imaging techniques produce images at a high-frame rate, streaming out a tremendous amount of image frames. This is not surprising, as phenomena occurring at a very small length scale also tend to operate on a short time scale; for instance, the nanometer scale transient behavior typically occurs in the scale of microseconds (10^{-6} s) to milliseconds (10^{-3} s). A single output image from an EM may have 2000×2000 image pixels, which amounts to 16 MB with 4 bytes per pixel. If the imaging rate is microseconds, the amount of image data being generated would be 16 TB (=16 MB × million images) per second, or petabytes (10^{15} bytes) per hour.

Analysis of dynamic images is at the frontier of material image analysis because of the relatively recent availability of the *in situ* instruments. Unsurprisingly, much fewer research studies have been reported on this topic.[45–49] It is an area for which data science and machine learning can make greater impacts, as the sheer amount of the data and the speed of its arrival make the manual analysis entirely out of the question.[50] The existing literature can be grouped into two categories: distribution tracking versus object tracking, depending on whether the interest is to track individual material objects and/or whether the material objects are traceable.

In some applications, the material objects may not be easily traceable. For example, in liquid phase EM experiments with a flow-through setup, samples of materials are continuously pumped into a TEM using microfluidics to flow through the imaging area, so that material samples observed at different times are different materials. Tracking individual material objects is thus infeasible. In some other applications, material scientists may care more about the collective change of material samples rather than the changes in individual objects. Under these circumstances, the collective behavior, or the growth trajectory of the group, can be represented using a probability density distribution. The dynamics to be tracked is the temporal evolution of a time-varying probability density distribution. We refer to this approach as distribution tracking.[45,46,48,49]

From the image analysis perspective, distribution tracking comes in two steps. The first step is to extract the outlines of material objects using the morphology analysis described in an earlier section of this paper, and the second step is to fit a probability distribution function or a probability density function to the characteristics of interest, e.g., size or shape. The major analytical challenge is in this second step, which is concerned with how the corresponding distribution function and its dynamics are modeled. One of the pioneering studies for modeling the evolution of structures was the introduction of the outline expansion model that explains the growth of material structures in time.[45,46] As illustrated in Fig. 4, a material structure is described by its exterior outline [Fig. 4(a)] or a curve representation [Fig. 4(b)]. The distribution of material structures is modeled as the variation of curves [Fig. 4(c)], which is represented by a nonparametric distribution of curves. Accordingly, the distributional change due to the growth of material structures is modeled as the time-varying coefficients

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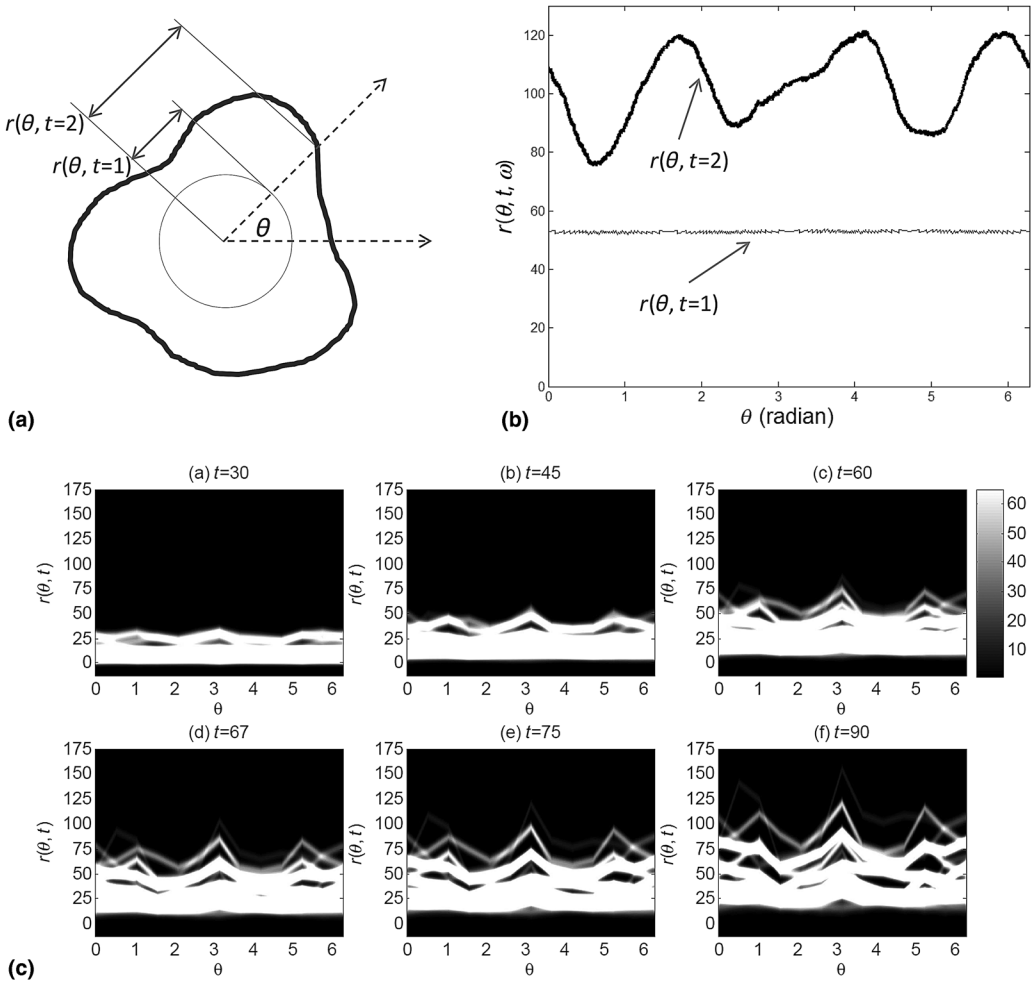


Figure 4. Analysis for shape and size evolution of an ensemble of nanoparticles: (a) an outline of material structures in 2d, (b) a curve representation of the outlines, and (c) the variation of material structures represented in terms of the variations of curves and the variations evolve in time. *Reprinted with permission from Park.[45]

in a nonparametric probability distribution. Related statistical inference and hypothesis testing procedures on the new model are also developed.[46] Dynamic material imaging calls for sophisticated development in an important data science branch, i.e., the nonparametric modeling of time-varying probability density functions, which is relatively underdevelopment,[51,52] due to both its challenging nature and the lack of real applications in the past.

The generality of the outline expansion model[45] comes with computationally expensive statistical inference steps. These computational issues have been addressed by adopting a new distribution representation.[48,49] In the new approach, the probability distribution is represented using a spline model, and the distributional changes were represented by the time-varying coefficients of the spline model. Estimating the spline model is much more computationally feasible, so that the resulting approach can be applied to larger datasets. The approach is employed to handle both retrospective and prospective analysis. The retrospective analysis[49] centers on off-line EM video data, to signal possible change points delineating the stages of growth; doing so produces strong clues about where to concentrate one's effort to understand the highly stochastic material evolving processes and enable discovery of the unexpected. The prospective analysis[48] is the real-time tracking, i.e., developing a dynamic and forward-looking model that can track the growth trajectory of a material characteristic, anticipate an upcoming change, and allow the design of specific interventions, to steer the course of a material evolution toward the desired target through the "on the fly" changes of process variables. Both the nonparametric approach[45,46] and spline approach[48,49] have their respective limitations. The spline approach may not be extendable to a high-dimensional representation of material structures. The nonparametric approach is more general but only applicable when the material structures can be abstracted by their exterior outlines. Future research on modern shape modeling and topological analysis is needed to extend the applicability of the existing studies to more complex material structures.

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Unlike in distribution tracking, the approach of object tracking is to specifically track individual material objects or structures over different image frames. Object tracking can be further classified into single-target tracking analysis and multi-target tracking analysis, depending on the number of material structures being tracked. Single-target tracking analysis is interested in how a single material object or structure evolves through interactions with chemical or biologic environment,[50] whereas multi-target tracking analysis is more interested in how material structures or objects evolve through mutual interactions among themselves as well as interactions with the external environment. Object tracking analysis requires one more step in addition to the morphology analysis of individual images. Once the morphology of materials is extracted from individual image frames (Fig. 5, step 1), it is associated with the same morphology over different image frames in order to form the trajectories of a material change (Fig. 5, step 2). For the single-target tracking analysis, the existing approaches in computer vision can be employed to extract the trajectory of a single material and its structure.[53–55] The existing approaches, however, are ineffective for the purpose of multi-target tracking analysis, because most of them are based on the assumption that there is no interaction among the objects being tracked. To material scientists, tracking mutual interactions among materials is of crucial importance. Only a few papers address the problem of tracking object interactions and do so to a limited degree, e.g., split and merge of two objects.[56,57]

A novel recent research is proposed for tracking complex interactions,[47] which is to formulate a multi-way data association problem that combines individual morphology data into trajectories of materials and material interactions, and encodes the interactions as a graph topology. Through the association, many imagery artifacts in the morphology data are filtered out, and miss-detected morphology is recovered, which ensures a better accuracy in tracking outcomes. Tracking the dynamic evolution of materials and material interactions would produce many material evolution trajectories in terms of changes in shape or structure. The number would be too numerous for manual scanning to pinpoint important findings. Statistical methods that are capable of summarizing numerous shape trajectories into a few frequent or rare trajectories are needed. Such methods, if developed, could significantly expedite the analysis of raw data and hypothesizing of the frequent and rare patterns (Fig. 5, step 3). Combining the tracking analysis and the subsequent pattern discovery would provide the kinetic rates of different material interactions and relate them to the overall material changes. The new multi-target tracking analysis has been successfully applied to many materials science problems, including nanocrystal growth,[10,58,59] *operando* analysis of the next generation battery system,[4,60] formation of metal organic frameworks,[11] and micelle evolutions.[61]

Compressive sensing and adaptive imaging

Machine learning and data science play an important role in improving the spatial or temporal resolutions of material imaging instruments through either optimization for higher resolutions or fusion of complementary data sources of different resolutions or those from instruments working at different scales.

One case in point is that the temporal resolution of STEM imaging is improved based on the compressive and adaptive sensing techniques. Compressive imaging or image subsampling is a novel approach in image acquisition. Its goal is to minimize the number of measurements needed to reconstruct an image with a prescribed accuracy. Compressive imaging is based on the principle of "sparsity," i.e., representing an image by using much fewer (i.e., sparser) bases in a proper basis set. In electron microscopy, compressive imaging has been applied to STEM imaging for reducing electron doses while imaging beam-sensitive materials such as organic and porous materials[62] and for accelerating the imaging speed when encountering a fast changing material process.[63,64] All existing approaches are based on random selection of measurement locations followed by an image in-painting reconstruction of the random measurements. The approaches are rather successful in terms of reconstructing atomic resolution images, partially for the reason that atom locations are sparse and well aligned to regular lattice coordinates, so that the sparsity and repeated patterns of the signals form an ideal condition for which the sparse random sampling works out greatly. However, when such a random and sparse sampling mechanism was applied to other electron microscopic imaging, the resulting reconstruction could lose sharp details over the interfacial areas of different materials, creating intolerable image blurriness. In those cases, it would be better to adaptively select measurement locations to improve the resulting image reconstruction.

In machine learning, a general approach for measurement selection to improve the accuracy of learning is called active learning,[65] also known as adaptive sampling or sequential experimental designs in engineering.[66] The overall approach for active learning is a multi-stage approach that iterates measurement taking and image reconstruction over time, making use of the image reconstructed during the past steps to guide the measurement taking process for the next step.[67] A major challenge in applying the active learning strategy in electron microscopy is caused by the repeated image reconstruction steps across numerous stages. Each image reconstruction could be computationally intensive and slow. Unsurprisingly, the repeated reconstructions slow down substantially the whole imaging process, which may cancel out the main advantage of using compressive imaging. Fast image construction with sparse samples should be developed before analysts can make full use of compressive sensing.

Closing remark and discussions

Material imaging is one of the major experimental tools enabling new material designs and discoveries. Different from other forms of data obtained by their respective

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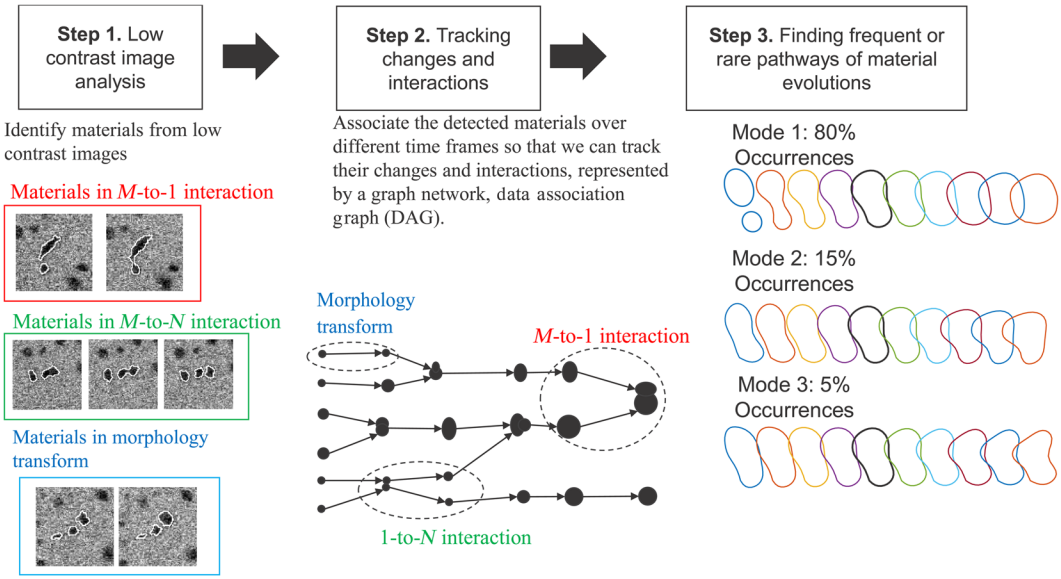


Figure 5. Three steps in multi-target tracking analysis of material evolutions and interactions. Step 1 detects individual material objects or structures in noisy images. Step 2 associates the detections from different image frames with each other for estimating the trajectories of material evolutions and interactions. Step 3 finds frequent and rare patterns in the trajectories.

characterization instruments, material image data is large in volume, contains complex data patterns in 2d and 3d, and starts to arrive in an overwhelming speed. The sheer volume and intrinsic complexity associated with material image data make material imaging a perfect big data candidate, calling for sophisticated data science and machine learning methodology development, so that the technology innovation in imaging instruments can be fully taken advantage of.

As reviewed and summarized in the paper, the recent developments have made remarkable inroad toward making material imaging methods automated, efficient, and potentially compatible with scaled-up production, especially in the following aspects: material morphology analysis, finding spatial symmetries and boundaries of materials, tracking dynamic material evolutions and interactions for material discovery, and improving temporal and spatial resolutions of imaging. These capabilities have already shown proven impacts on materials research, providing material characterization and information from otherwise unanalyzable sizes of scientific image data.

We would like to stress that all these efforts are the starting point in addressing the continuing challenges and practical needs in material image analysis. We summarize several ongoing research issues, based on our experience and interactions with domain experts, in the following aspects:

- *On-the-fly data reduction or compressive sensing*. The current practice of analyzing scientific imaging data first stores the data at a local data storage site and then performs subsequent analysis. This standard technique is time consuming, mainly due to slow disk writing and data communication speed. For example, the state-of-the-art in disk I/O interface has a writing speed of several GBs per second. This means that it would take 4000 s, or about 1 h, to write into storage the 4 TB *in situ* electron microscopic data, which could have been generated for a single second of a nanometer scale process; or equivalently, it would take 3600 h (150 days) to write the whole data set generated for an hour of the same process. Due to the slow disk I/O speed, in relative to the much faster data generation rate, data reduction on-the-fly during imaging is highly desirable and necessary.
- *Enhancement of computational efficiency*. This is a fundamental problem applicable to almost every aspect of material image analysis. With the increasing number of material objects or structures and the increasing complexity associated with each of them, making the models/algorithms run fast is not a trivial undertaking. Yet, a fast algorithm is absolutely essential leading to practical adoption of both offline and online methods. The online need is self-evident, as the time pressure is dictated by the process dynamics. Even for offline analysis, analysts or scientists are unlikely to adopt an algorithm that takes too long to produce an outcome. The current state-of-the-art in image binarization and segmentation for morphology analysis takes a few seconds to minutes to analyze a typical SEM or TEM image, and the tracking analysis could take a longer time; all these are much slower than the image generation speed. Without cutting the analysis time close to the image generation speed, data storage could quickly explode with a large amount of unanalyzed images. To accelerate the analysis to the order of milliseconds or microseconds, at par with the image generation speed, a combination of new algorithmic developments and new hardware acceleration of the algorithms appears to be inevitable.

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- *From 2d to 3d*. We anticipate that the main focus in material imaging would soon shift from 2d imaging toward 3d imaging and even from 3d imaging to 4d imaging (3d spatial dimensions plus a temporal dimension).[68] The machine learning and data science approaches reviewed earlier serve as a good starting point to automate the data analysis for 3d imaging, but future research must follow to handle the ever-increasing volume and complexity. One foundational problem is the *effective shape and topology representations of material structures in 3d*. Most of the current shape data analyses are focused on planar shapes in 2d, often modeled with various kinds of curve representations. This type of models cannot describe complex material structures and topology in 3d. Mesh-based approaches in computational geometry is a natural extension of the curve representation to 3d structures and could be potentially useful for constructing a new shape space in 3d, but developing the notions of invariances and similarity metrics as well as the corresponding statistical estimation in the new space is rather difficult. Moreover, the mesh approaches may not be efficient, because they require a large degree of freedom to represent a geometry in 3d, creating a high-dimensional feature space, hard to handle in modeling and solution steps. Proper dimension reduction and feature extraction are necessary.
- *Super-resolution*. Related to the discussion in the section of adaptive sampling, an image processing technique, known as super-resolution, is being introduced to material image processing.[69] The general concept of super-resolution refers to the class of methods that enhance the resolution of an imaging system, but here we refer more specifically to the enhancements resulting from the algorithmic or image processing aspect. For material imaging problems, with the presence of multi-resolution electron microscopic images, it saves a lot of time and efforts if one can boost the quality of images taken by a low-resolution microscope through the use of a small fraction of high-resolution data. This problem is apparently connected with a set of statistical learning problems, originated in the field of computer experiments, in which physical experiment data and computer simulation data, or data coming from multi-fidelity computer models, are combined for making better predictions or inference at settings or conditions where the high-fidelity or high-quality response data are not available.[70–72] Making intelligent use of multi-resolution data sources for achieving the super-resolution objective may help address some of the efficiency and complexity issues raised above.
- *Process and quality control*. The availability of *in situ* electron microscopy gives the hopes to real-time process control. The goal of process control is to take process variable conditions as inputs, builds predictive models connecting the controllable settings in a material exploration process or in a manufacturing process to the intermediate, as well as the final, quality characteristics, and recommends appropriate process adjustments needed for achieving the quality control targets. The dynamic imaging models, especially the real-time tracking and updating capabilities as discussed in the paper, lay the critical corner stone for realizing the process control objective. But in order to fully materialize process control, there are still plenty of questions left unanswered, including the predictability and controllability of underlying material processes and the stability and robustness of the control.

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References

1. *Basic Research Needs for Innovation and Discovery of Transformative Experimental Tools*; available at http://science.energy.gov, 2017.
2. A.V. Crewe: Scanning transmission electron microscopy. *J. Microsc.* **100**, 247–259 (1974).
3. S.M. Salapaka and M.V. Salapaka: Scanning probe microscopy. *IEEE Control Syst.* **28**, 65–83 (2008).
4. P. Abellan, B.L. Mehdi, L.R. Parent, M. Gu, C. Park, W. Xu, Y. Zhang, I. Arslan, J.-G. Zhang, and C.-M. Wang: Probing the degradation mechanisms in electrolyte solutions for Li-ion batteries by *in situ* transmission electron microscopy. *Nano Lett.* **14**, 1293–1299 (2014).
5. M.P. Chien, M.P. Thompson, C.V. Barback, T.H. Ku, D.J. Hall, and N.C. Gianneschi: Enzyme-directed assembly of a nanoparticle probe in tumor tissue. *Adv. Mater.* **25**, 3599–3604 (2013).
6. J.E. Evans, K.L. Jungjohann, N.D. Browning, and I. Arslan: Controlled growth of nanoparticles from solution with *in situ* liquid transmission electron microscopy. *Nano Lett.* **11**, 2809–2813 (2011).
7. J.S. Kim, T. LaGrange, B.W. Reed, M.L. Taheri, M.R. Armstrong, W.E. King, N.D. Browning, and G.H. Campbell: Imaging of transient structures using nanosecond *in situ* TEM. *Science* **321**, 1472–1475 (2008).
8. T. LaGrange, G.H. Campbell, B. Reed, M. Taheri, J.B. Pesavento, J.S. Kim, and N.D. Browning: Nanosecond time-resolved investigations using the *in situ* of dynamic transmission electron microscope (DTEM). *Ultramicroscopy* **108**, 1441–1449 (2008).
9. T.J. Woehl, J.E. Evans, I. Arslan, W.D. Ristenpart, and N.D. Browning: Direct *in situ* determination of the mechanisms controlling nanoparticle nucleation and growth. *ACS Nano* **6**, 8599–8610 (2012).
10. T.J. Woehl, C. Park, J.E. Evans, I. Arslan, W.D. Ristenpart, and N.D. Browning: Direct observation of aggregative nanoparticle growth: kinetic modeling of the size distribution and growth rate. *Nano Lett.* **14**, 373–378 (2013).
11. J. P. Patterson, P. Abellan, M. S. Denny Jr., C. Park, N. D. Browning, S. M. Cohen, J. E. Evans, and N. C. Gianneschi: Observing the growth of metal–organic frameworks by *in situ* liquid cell transmission electron microscopy. *J. Am. Chem. Soc.* **137**, 7322–7328 (2015).
12. S. Jesse and S.V. Kalinin: Band excitation in scanning probe microscopy: sines of change. *J. Phys. D: Appl. Phys.* **44**, 464006 (2011).
13. B.J. Rodriguez, C. Callahan, S.V. Kalinin, and R. Proksch: Dual-frequency resonance-tracking atomic force microscopy. *Nanotechnology* **18**, 475504 (2007).
14. S.V. Kalinin, E. Strelcov, A. Belianinov, S. Somnath, R.K. Vasudevan, E.J. Lingerfelt, R.K. Archibald, C. Chen, R. Proksch, N. Laanait, and S. Jesse: Big, deep, and smart data in scanning probe microscopy. *ACS Nano* **10**, 9068–9086 (2016).
15. M.C. Roco: The Long View of Nanotechnology Development: The National Nanotechnology Initiative at 10 Years. *Journal of Nanoparticles* **13**, 427–445 (2011).
16. J. Canny: A computational approach to edge detection. *IEEE Trans. Pattern Anal. Mach. Intell.* **8**, 679–698 (1986).

https://doi.org/10.1557/mrc.2019.48 Published online by Cambridge University Press

17. N. Otsu: A threshold selection method from gray-level histograms. *IEEE Trans. Syst. Man Cybern.* **9**, 62–66 (1979).
18. X. Jiang and D. Mojon: Adaptive local thresholding by verification-based multithreshold probing with application to vessel detection in retinal images. *IEEE Trans. Pattern Anal. Mach. Intell.* **25**, 131–137 (2003).
19. G. Vo and C. Park: Robust regression for image binarization under heavy noises and nonuniform background. *Pattern Recognit.* **81**, 224–239 (2018).
20. C. Park, J.Z. Huang, D. Huitink, S. Kundu, B.K. Mallick, H. Liang, and Y. Ding: A multistage, semi-automated procedure for analyzing the morphology of nanoparticles. *IIE Trans.* **44**, 507–522 (2012).
21. C. Park, J.Z. Huang, J.X. Ji, and Y. Ding: Segmentation, inference and classification of partially overlapping nanoparticles. *IEEE Trans. Pattern Anal. Mach. Intell.* **35**, 669–681 (2013).
22. S. Beucher and F. Meyer: The morphological approach to segmentation: the watershed transformation. *Optical Engineering* **34**, 433–433 (1992).
23. Y. Qian, J.Z. Huang, X. Li, and Y. Ding: Robust nanoparticles detection from noisy background by fusing complementary image information. *IEEE Trans. Image Process.* **25**, 5713–5726 (2016).
24. S. Zafari, T. Eerola, J. Sampo, H. Kälviäinen, and H. Haario: Segmentation of overlapping elliptical objects in silhouette images. *IEEE Trans. Image Process.* **24**, 5942–5952 (2015).
25. X.-L. Meng, and D.B. Rubin: Maximum likelihood estimation via the ECM algorithm: a general framework. *Biometrika* **80**, 267–278 (1993).
26. P. Bubenik: Statistical topological data analysis using persistence landscapes. *J. Mach. Learn. Res.* **16**, 77–102 (2015).
27. B.A. Konomi, S.S. Dhavala, J.Z. Huang, S. Kundu, D. Huitink, H. Liang, Y. Ding, and B.K. Mallick: Bayesian object classification of gold nanoparticles. *Ann. Appl. Stat.* **7**, 640–668 (2013).
28. J. Frank: *Electron Tomography: Three-Dimensional Imaging with the Transmission Electron Microscope* (New York, NY: Springer Science & Business Media, 2013).
29. C. Mu and C. Park: Optimal filtered backprojection for fast and accurate tomography reconstruction. *Pattern Recognition* Submitted (2019).
30. X. Li, A. Belianinov, O. Dyck, S. Jesse, and C. Park: Two-level structural sparsity regularization for identifying lattices and defects in noisy images. *Ann. Appl. Stat.* **12**, 348–377 (2018).
31. L. Dong, X. Li, Y. Qian, D. Yu, H. Zhang, Z. Zhang, and Y. Ding: Quantifying nanoparticle mixing state to account for both location and size effects. *Technometrics* **59**, 391–403 (2017).
32. A. Belianinov, Q. He, M. Kravchenko, S. Jesse, A. Borisevich, and S.V. Kalinin: Identification of phases, symmetries and defects through local crystallography. *Nat. Commun.* **6**, 7801 (2015).
33. D.S. Bright and E.B. Steel: Two-dimensional top hat filter for extracting spots and spheres from digital images. *J. Microsc.* **146**, 191–200 (1987).
34. D. Sage, F.R. Neumann, F. Hediger, S.M. Gasser, and M. Unser: Automatic tracking of individual fluorescence particles: application to the study of chromosome dynamics. *IEEE Trans. Image Process.* **14**, 1372–1383 (2005).
35. J. Hughes, J. Fricks, and W. Hancock: Likelihood inference for particle location in fluorescence microscopy. *Ann. Appl. Stat.* **4**, 830–848 (2010).
36. N. Laanait, M. Ziatdinov, Q. He, and A. Borisevich: Identifying local structural states in atomic imaging by computer vision. *Adv. Struct. Chem. Imaging* **2**, 14 (2017).
37. B.D. Ripley: The second-order analysis of stationary point processes. *J. Appl. Probab.* **13**, 255–266 (1976).
38. X. Li, H. Zhang, J. Jin, D. Huang, X. Qi, Z. Zhang, and D. Yu: Quantifying dispersion of nanoparticles in polymer nanocomposites through transmission electron microscopy micrographs. *J. Micro Nano-Manufacturing* **2**, 021008 (2014).
39. N. De Jonge and F.M. Ross: Electron microscopy of specimens in liquid. *Nat. Nanotechnol.* **6**, 695 (2011).
40. S.V. Kalinin, B.G. Sumpter, and R.K. Archibald: Big-deep-smart data in imaging for guiding materials design. *Nat. Mater.* **14**, 973 (2015).
41. H. Zheng, Y.S. Meng, and Y. Zhu: Frontiers of *in situ* electron microscopy. *MRS Bull.* **40**, 12–18 (2015).
42. M. Grzelczak, J. Vermant, E.M. Furst, and L.M. Liz-Marzán: Directed self-assembly of nanoparticles. *ACS Nano* **4**, 3591–3605 (2010).
43. A. Mikhailov and G. Gundersen: Relationship between microtubule dynamics and lamellipodium formation revealed by direct imaging of microtubules in cells treated with nocodazole or taxol. *Cytoskeleton* **41**, 325–340 (1998).
44. L.G. Bergen and G.G. Borisy: Head-to-tail polymerization of microtubules in vitro. Electron microscope analysis of seeded assembly. *J. Cell Biol.* **84**, 141–150 (1980).
45. C. Park: Estimating multiple pathways of object growth using nonlongitudinal image data. *Technometrics.* **56**, 186–199 (2014).
46. C. Park and A.K. Shrivastava: Multimode geometric-profile monitoring with correlated image data and its application to nanoparticle self-assembly processes. *J. Qual. Technol.* **46**, 216–233 (2014).
47. C. Park, T.J. Woehl, J.E. Evans, and N.D. Browning: Minimum cost multi-way data association for optimizing multitarget tracking of interacting objects. *IEEE Trans. Pattern Anal. Mach. Intell.* **37**, 611–624 (2015).
48. Y. Qian, J. Z. Huang; C. Park, and Y. Ding: Fast dynamic nonparametric distribution tracking in electron microscopic data. *Ann. Appl. Stat.* in press (2019).
49. Y. Qian, J.Z. Huang, and Y. Ding: Identifying multi-stage nanocrystal growth using *in situ* TEM video data. *IISE Trans.* **49**, 532–543 (2017).
50. H. Zheng, R.K. Smith, Y.-W. Jun, C. Kisielowski, U. Dahmen, and A.P. Alivisatos: Observation of single colloidal platinum nanocrystal growth trajectories. *Science* **324**, 1309–1312 (2009).
51. A. Rodriguez and E. Ter Horst: Bayesian dynamic density estimation. *Bayesian Anal.* **3**, 339–365 (2008).
52. R.H. Mena and M. Ruggiero: Dynamic density estimation with diffusive Dirichlet mixtures. *Bernoulli. (Andover)* **22**, 901–926 (2016).
53. H. Jiang, S. Fels, and J. J. Little: In *A linear programming approach for multiple object tracking*, 2007 IEEE Conference on Computer Vision and Pattern Recognition, IEEE: 2007; pp. 1–8.
54. K. Okuma, A. Taleghani, N. De Freitas, J. J. Little, and D. G. Lowe: In *A boosted particle filter: Multitarget detection and tracking*, 2004 European Conference on Computer Vision, Springer, 2004; pp. 28–39.
55. H. Pirsiavash, D. Ramanan, and C. C. Fowlkes: In Globally-optimal greedy algorithms for tracking a variable number of objects, 2011 IEEE Conference on Computer Vision and Pattern Recognition, IEEE, 2011; pp. 1201–1208.
56. K. Jaqaman, D. Loerke, M. Mettlen, H. Kuwata, S. Grinstein, S.L. Schmid, and G. Danuser: Robust single-particle tracking in live-cell time-lapse sequences. *Nat. Methods* **5**, 695 (2008).
57. J. F. Henriques, R. Caseiro, and J. Batista: In *Globally optimal solution to multi-object tracking with merged measurements*, 2011 IEEE International Conference on Computer Vision, IEEE, 2011; pp. 2470–2477.
58. D.A. Welch, T.J. Woehl, C. Park, R. Faller, J.E. Evans, and N.D. Browning: Understanding the role of solvation forces on the preferential attachment of nanoparticles in liquid. *ACS Nano* **10**, 181–187 (2015).
59. A. Esmaieeli Sikaroudi, D.A. Welch, T.J. Woehl, R. Faller, J.E. Evans, N.D. Browning, and C. Park: Directional statistics of preferential orientations of two shapes in their aggregate and Its application to nanoparticle aggregation. *Technometrics* **60**, 332–344 (2018).
60. B.L. Mehdi, J. Qian, E. Nasybulin, C. Park, D.A. Welch, R. Faller, H. Mehta, W.A. Henderson, W. Xu, and C.M. Wang: Observation and quantification of nanoscale processes in lithium batteries by operando electrochemical (S) TEM. *Nano Lett.* **15**, 2168–2173 (2015).
61. M.A. Touve, C.A. Figg, D.B. Wright, C. Park, J. Cantlon, B.S. Sumerlin, and N.C. Gianneschi: Polymerization-induced self-assembly of micelles observed by liquid cell transmission electron microscopy. *ACS Cent. Sci.* **4**, 543–547 (2018).
62. A. Stevens, L. Luzi, H. Yang, L. Kovarik, B. Mehdi, A. Liyu, M. Gehm, and N. Browning: A sub-sampled approach to extremely low-dose STEM. *Appl. Phys. Lett.* **112**, 043104 (2018).
63. L. Kovarik, A. Stevens, A. Liyu, and N.D. Browning: Implementing an accurate and rapid sparse sampling approach for low-dose atomic resolution STEM imaging. *Appl. Phys. Lett.* **109**, 164102 (2016).
64. A. Stevens, L. Kovarik, P. Abellan, X. Yuan, L. Carin, and N.D. Browning: Applying compressive sensing to TEM video: a substantial frame rate increase on any camera. *Adv. Struct. Chem. Imaging* **1**, 10 (2015).

https://doi.org/10.1557/mrc.2019.48 Published online by Cambridge University Press

65. R. Castro, J. Haupt, and R. Nowak: In *Compressed sensing vs. active learning*, 2006 IEEE International Conference on Acoustics, Speech and Signal Processing, IEEE, 2006; p. III.
66. R. Edgeworth and R.G. Wilhelm: Adaptive sampling for coordinate metrology. *Prec. Eng.* **23**, 144–154 (1999).
67. C. Park and P. Qiu: Sequential Adaptive Design for Jump Regression Estimation. Submitted (IEEE Transactions on Pattern Analysis and Machine Intelligence 2019). Also available at https://arxiv.org/abs/1904.01648
68. A. H. Zewail and J. M. Thomas: *4D Electron Microscopy: Imaging in Space and Time*. (Imperial College Press: London, 2009).
69. S. Sreehari, S. Venkatakrishnan, K. L. Bouman, J. P. Simmons, L. F. Drummy, and C. A. Bouman: In *Multi-resolution data fusion for super-resolution electron microscopy*, 2017 IEEE Conference on Computer Vision and Pattern Recognition Workshops, 2017; pp. 1084–1092.
70. H. Xia, Y. Ding, and B.K. Mallick: Bayesian hierarchical model for combining misaligned two-resolution metrology data. *IIE Trans.* **43**, 242–258 (2011).
71. A.A. Ezzat, A. Pourhabib, and Y. Ding: Sequential design for functional calibration of computer models. *Technometrics*. **60**, 286–296 (2018).
72. A. Pourhabib, J.Z. Huang, K. Wang, C. Zhang, B. Wang, and Y. Ding: Modulus prediction of buckypaper based on multi-fidelity analysis involving latent variables. *IIE Trans.* **47**, 141–152 (2015).

https://doi.org/10.1557/mrc.2019.48 Published online by Cambridge University Press