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Predicting colostrum and calf blood components based on refractometry

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Research Article

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Abstract

Provision of good quality colostrum is essential for the passive immunity and nutrition of newborn calves. In order to better predict the quality of colostrum and the transfer of passive immunity, the relationships between colostrum components and between calf serum components were examined in this study. Samples of bulk tank milk, colostrum pooled from several cows 0–4 d postpartum, and colostrum collected from individual cows twice daily for 3 d postpartum were compared. With the exception of fat percentage, there were strong correlations between the levels of the components in the pooled colostrum and in the individual cow colostrum collected 0–1 d postpartum. The correlations between total solids as measured by Brix refractometry and total protein, immunoglobulin G (IgG), lactose % and protein % in colostrum within 1 d postpartum and pooled colostrum were 0.92, 0.90, −0.88 and 0.98, respectively. These high correlations enabled these colostrum components to be accurately predicted from Brix % and therefore, the volume of colostrum required to feed neonate calves can be optimised based on Brix refractometry to avoid failure of passive immunity transfer. To assess whether the components obtained from colostrum were correlated in calf blood, newborn calves were separated from their dams before suckling and blood sampled before feeding (day 0), and on days 1 and 7, after receiving colostrum or milk twice a day. The correlations between glucose, total protein, IgG, and gamma-glutamyl transferase (GGT) levels in the calf blood were lower than the correlations observed between the colostrum components. The highest correlation was between serum protein measured by refractometer and serum IgG within one week postpartum. GGT activity was not a good indicator of serum IgG levels. However, serum protein refractometer measurements predicted serum IgG level with high accuracy, providing an on-farm test to determine that calves have received sufficient passive immunity and colostrum components.

The morphology of bovine placenta prevents the transfer of immunoglobulins from dam to foetus during pregnancy, therefore, colostrum is essential for the transfer of passive immunity to newborn calves (Barrington and Parish, 2001; Castro *et al.*, 2011). Failure of passive immunity transfer (FPIT) may occur if a calf does not receive enough good-quality colostrum soon after birth and absorb sufficient immunoglobulins, specifically immunoglobulin G (IgG). Failure of passive immunity transfer is associated with increased calf morbidity and mortality (Barrington and Parish, 2001; Moran, 2002; Furman-Fratczak *et al.*, 2011; Vandeputte *et al.*, 2011), and avoiding FPIT is crucial to optimise calf health and productivity.

Failure of passive immunity transfer can be detected in calves by measuring serum IgG or serum protein or by estimating the total serum protein using refractometry. The generally accepted FPIT thresholds at 24 h postpartum are less than 10 g/l of serum IgG (Weaver *et al.*, 2000; Godden *et al.*, 2019; Oliveira *et al.*, 2019) or less than 52 g/l of serum total protein (TP) (McGuirk and Collins, 2004; Hernandez *et al.*, 2016; Cuttance *et al.*, 2017*b*). Other authors have estimated a Brix refractometry threshold for FPIT as less than 8.1–8.8% for calf serum at 24–48 h postpartum (Hernandez *et al.*, 2016; Cuttance *et al.*, 2017*a*; Godden *et al.*, 2019). It has been suggested that FPIT can be also inferred by the gamma-glutamyl transferase (GGT) levels in the calf blood with levels less than 200 IU/l of GGT at 24 h postpartum indicating FPIT (Perino *et al.*, 1993; Parish *et al.*, 1997).

To avoid FPIT, it is recommended that calves receive approximately 10% of their body weight of good quality colostrum (>50 g/l of IgG) within few hours of birth (McGuirk and Collins, 2004; Bartier *et al.*, 2015; Godden *et al.*, 2019). By quantifying the components in the colostrum before feeding the calf, the quality can be assessed so that a sufficient volume can be given to avoid FPIT. There are many laboratory-based methods for quantifying IgG to determine colostrum quality (Gapper *et al.*, 2007). However, these laboratory-based methods are time consuming, costly, and difficult to apply routinely as samples need to be transported to the laboratory.

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On-farm techniques for estimating the IgG concentration in samples that provide results rapidly can be used to manage the colostrum fed to neonatal calves and to monitor the transfer of colostrum components to the calves. Two such techniques are Brix refractometry for colostrum samples (Quigley *et al.*, 2013; Morrill *et al.*, 2015; Cabral *et al.*, 2016) and digital serum refractometry for blood samples (Vandeputte *et al.*, 2011; Deelen *et al.*, 2014; Hernandez *et al.*, 2016). Brix refractometry values between 18 and 23% are considered to be equivalent to 50 g/l IgG, the threshold for good quality colostrum (Bielmann *et al.*, 2010; Quigley *et al.*, 2013; Bartier *et al.*, 2015; Morrill *et al.*, 2015).

In addition to providing passive immunity, colostrum contains all the nutrients that are essential for the survival of the neonate calf, including proteins, lactose, fat, minerals, and vitamins (McGrath *et al.*, 2016; Godden *et al.*, 2019). However, colostrum quality is rarely considered in terms of these other components, and their assessment on-farm has not been studied. In the present study, the components of colostrum, milk and calf blood were quantified by both refractometry and laboratory assays and their correlations determined. The data were then used to develop linear regression models to determine the predictability of colostrum component levels or blood component levels based on refractometry measurements or the other components.

Materials and methods

Sample collection

The animal work was performed at a commercial dairy farm with over 1800 cows located in Mount Gambier, South Australia. All animal experimental work was approved by the University of Adelaide Animal Ethics Committee (approval number S-2017-060). Twelve Holstein-Friesian cows were milked within 2 h postpartum prior to being suckled (day 0), and then twice daily for 3 d (days 1, 2 and 3). The colostrum and transition milk samples were collected every 24 h during this time and are referred to as 'individual cow colostrum'. Twenty five (25) samples of bulk tank milk were also collected. The 'bulk tank milk' was from the all cows on the dairy farm at 5 d or more postpartum. In addition, six samples were collected from the pooled colostrum and transitional milk of the other cows on the farm at 0–4 d postpartum. These samples are referred to as 'pooled colostrum' to distinguish from the 'bulk tank milk' and 'individual cow colostrum' samples. Total solids in the colostrum and milk samples were measured immediately after collection by Brix refractometry (digital refractometer DBR-1, Star Instrument with a measurement range from Brix 0–50%). The colostrum and milk samples were aliquoted into sterile 50 ml-tubes, frozen at −20°C, and stored at −80°C until analysed.

Thirty-five Holstein-Friesian bull calves born in February and March 2018 were divided into 2 groups using block randomisation by birth order. The first group ($n = 24$) was fed colostrum from the individual cows (5% of body birth weight/feed) harvested at the milking just prior to feeding twice a day for 3 d, and then fed bulk tank milk for 4 d. The second group of calves ($n = 11$) was fed one bottle of pooled colostrum at the first feed (1.7 litres), and then fed bulk tank milk twice a day for 7 d.

Blood samples were collected into 6 ml – BD vacutainers without anticoagulant and BD vacutainers with heparin from all calves within 4 h after birth before their first feed (day 0) and at day 1 and day 7 postpartum. Glucose concentration in whole calf blood was measured using a glucose meter immediately after collection (Accu-check Performa, Roche Diabetes Care, Basel, Switzerland). The blood was then separated immediately by centrifugation (2000 ***g***) for 15 min at room temperature to obtain serum and plasma. The total protein was measured in serum immediately after centrifugation using a serum protein refractometer (TP-R) (ATAGO Digital Refractometer with a protein measurement range from 0 to 12 g/100 ml). The serum and plasma samples were frozen at −20°C and stored at −80°C until analysed.

Colostrum and calf blood component measurements

The IgG concentration in the individual cow colostrum, pooled colostrum, bulk tank milk samples and calf serum samples was quantified by enzyme-linked immunosorbent assay (ELISA) using two bovine IgG specific antibodies (Life Technologies, USA) as described in the Supplementary File. Total protein in the individual cow colostrum, pooled colostrum, bulk tank milk and calf serum samples was assayed in a 96 well-plate using a Quick Start Bradford Protein assay kit (TP-B) (Bio-Rad Laboratories, Inc., USA) following the manufacturer's instructions as detailed in the Supplementary File.

Percentages of total protein, lactose and fat concentration in the thawed colostrum and milk samples were determined using a Fourier-transform mid-range infrared analyser (FT-MIR spectrometer, Foss Analytics) at the National Herd Development Co-operative (Kyabram VIC 3620, Australia). Calf plasma was analysed for gamma-glutamyl transferase (GGT) levels using a kinetic colour test with a Beckman Coulter analyser at the Veterinary Diagnostic Laboratory, the University of Adelaide, South Australia.

Statistical analysis

Statistical analyses were conducted in R (R version 3.6.3) and independently verified in SAS (Edition 9.4, SAS Institute, Cary, NC, USA). Descriptive statistics for the total solids, total protein, IgG, protein %, fat %, and lactose % in colostrum, and the glucose concentration, the total protein, IgG and GGT in calf blood were calculated and the correlations between the various components in the colostrum and milk samples and in the calf blood samples were determined. The correlations were categorised as follows: negligible ($r < 0.30$); low ($r = 0.30$ to 0.49); moderate ($r = 0.50$ to 0.69); high ($r = 0.70$ to 0.89); or very high ($r \geqslant 0.90$) (Wilm *et al.*, 2018). To evaluate the association between any two components, simple linear regression analysis was performed using linear models for two continuous variables. The accuracy of the predictions was estimated using the coefficient of determination (R^2). The coefficient of determination (R^2), or goodness of fit, has a value between 0.0 and 1.0, where a value of 1.0 indicates a perfect fit and a value of 0.0 indicates that the model fails to accurately predict the data.

Results and discussion

Concentration of colostrum components

The constituents of individual cow colostrum collected days 0–3 postpartum, pooled colostrum and bulk tank milk were compared. The pooled colostrum replicated the dairy's practice of mixing colostrum and transition milk from all cows 0 to 4 d postpartum to feed to the newborn calves.

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Table 1. Descriptive statistics of different colostrum and milk components

Sample	IgG (g/l)	Total solids (Brix %)	TP-B (g/l)	Protein (%)	Fat (%)	Lactose (%)
Cow day 0 ($n = 12$)						
Mean ± SE	184.4 ± 20.4	25.4 ± 1.4	170.8 ± 16.0	16.6 ± 0.9	4.6 ± 0.4	1.9 ± 0.2
Range	84.7–330.7	18.5–34.0	88.7–260.5	11.0–21.2	2.2–6.6	1.1–3.0
Cow day 1 ($n = 11$)						
Mean ± SE	29.4 ± 6.6	12.2 ± 0.6	51.2 ± 5.8	6.3 ± 0.5	3.2 ± 0.4	4.2 ± 0.1
Range	5.5–63.5	9.0–16.1	23.6–80.3	3.9–8.7	1.6–6.1	3.7–4.9
Cow day 2 ($n = 12$)						
Mean ± SE	4.9 ± 0.6	10.6 ± 0.2	34.4 ± 2.1	–	–	–
Range	2.3–9.4	8.8–12.2	19.1–44.7	–	–	–
Cow day 3 ($n = 12$)						
Mean ± SE	2.7 ± 0.6	10.5 ± 0.3	34.4 ± 2.1	–	–	–
Range	1.2–8.0	8.7–11.9	23.1–51.0	–	–	–
Pooled colostrum ($n = 6$)						
Mean ± SE	90.5 ± 9.6	19.4 ± 2.1	88.7 ± 11.6	12.4 ± 1.3	5.4 ± 0.9	2.4 ± 0.1
Range	69.7–131.6	12.3–25.9	54.3–127.8	–	–	–
Bulk tank milk ($n = 25$)						
Mean ± SE	0.3 ± 0.1	8.9 ± 0.2	28.6 ± 2.0	–	–	–
Range	0.2–0.7	7.6–12.0	7.1–50.1	–	–	–

IgG = immunoglobulin G; TP-B = total protein measured by Bradford assay; Cow = individual cow colostrum at the time point (day) indicated; Pooled colostrum = colostrum and transition milk pooled from cows days 0–4 postpartum; Bulk tank milk = milk collected from cows more than 4 d postpartum.

The IgG concentration in the individual cow colostrum decreased dramatically from a mean value of 184.4 g/l at day 0 to 2.7 g/l by day 3 (Table 1). The level of IgG in the pooled colostrum was relatively high (90.5 g/l), but was very low in the bulk tank milk (0.3 g/l). Both the individual cow colostrum and pooled colostrum had IgG concentrations that were higher than the threshold of 50 g/L recommended to avoid FPIT (McGuirk and Collins, 2004; Baumrucker *et al.*, 2014; Bartier *et al.*, 2015).

Immediately postpartum, the level of total solids in the individual cow colostrum was high (25.4 ± 1.4 Brix %), but declined rapidly by day 1 (12.2 ± 0.6 Brix %). The Brix % observed in all the individual cow colostrum samples immediately postpartum was higher than the published thresholds of 18–23% for good quality colostrum. This is presumably because the colostrum was obtained from the individual cows within 2 h postpartum and the cows were multiparous. The mean Brix % was also high in pooled colostrum (19.4 ± 2.1%), although two of the samples were below 18%, the Brix threshold for good quality colostrum (Morrill *et al.*, 2015).

Total protein in the individual cow colostrum, as measured by Bradford assay and FT-MIR, had the same trend as the levels of IgG and total solids, with very high levels on day 0 (170.8 ± 16.0 g/l, 16.6 ± 0.9%), a sharp decline by day 1 (51.2 ± 5.8 g/l, 6.3 ± 0.5%) and a constant level thereafter (34.4 ± 2.1 g/l at days 2 and 3). Total protein levels in the pooled colostrum were roughly half of the values of the individual cow colostrum between days 0 and 1 (88.7 ± 11.6 g/l, 12.4 ± 1.3%), and the level of total protein in the bulk tank milk was similar to the day 2 and day 3 individual cow colostrum (28.6 ± 2.0 g/l).

The fat concentration in the individual cow colostrum at day 0 was slightly lower than the pooled colostrum (4.6 ± 0.4% and 5.4 ± 0.9% respectively) and declined by day 1 (3.2 ± 0.4%). The decline in colostrum fat percentage one day postpartum was also observed by Dunn *et al.* (2017), who found the fat percent in colostrum decreased from 7.0% to 6.0% within 24 h postpartum and from 6.7% to 3.9% between the first and third milking postpartum (reviewed by Godden *et al.* (2019)).

In contrast to the other components, the lactose concentration in the individual cow colostrum increased from day 0 to day 1 (1.9 ± 0.2% and 4.2 ± 0.1%, respectively). The pooled colostrum was intermediate (2.4 ± 0.1%). These lactose levels were comparable to those obtained in other studies where lactose was found to be low in colostrum but to increase in the milk as lactation is established (Kehoe *et al.*, 2007).

Correlations between colostrum components

The correlations between components in the individual cow colostrum from days 0 and 1 postpartum were high (Table 2). The correlations between total solids measured by Brix refractometer with total protein, IgG, lactose % and protein % were 0.92, 0.90, −0.88 and 0.98, respectively, for the combined data from the pooled colostrum and the individual cow colostrum days 0 and 1 postpartum samples ($n = 29$) (Table 2).

Correlations between Brix refractometer measurements and colostrum IgG measured by radial immunodiffusion (RID) or turbidimetric immunoassay (TIA), previously reported for day 0 and 1 post-partum, range between 0.64 and 0.75 (Bielmann *et al.*, 2010; Quigley *et al.*, 2013; Bartier *et al.*, 2015; Elsohaby *et al.*, 2017). In the present study, the IgG concentration was measured by sandwich ELISA, which has higher sensitivity and specificity for IgG than other methods, including RID or TIA (Li-Chan

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Table 2. Correlations between components in colostrum and milk

Correlated parameters	Individual cow colostrum				Bulk tank milk (*n* = 25)	Cow + pooled colostrum (*n* = 29)
	Day 0 (*n* = 12)	Day 1 (*n* = 11)	Day 2 (*n* = 12)	Day 3 (*n* = 12)		
Total solids and TP-B	0.82	0.80	−0.02	0.50	0.15	0.92
Total solids and IgG	0.80	0.60	0	0.13	0.36	0.90
Total solids and fat %	0.62	0.08	[a]	[a]	[a]	0.54
Total solids and lactose %	−0.84	−0.71	[a]	[a]	[a]	−0.88
Total solids and protein %	0.94	0.90	[a]	[a]	[a]	0.98
TP-B and IgG	0.69	0.76	0.41	0.76	−0.13	0.90
TP-B and fat %	0.43	0.28	[a]	[a]	[a]	0.40
TP-B and lactose %	−0.66	−0.70	[a]	[a]	[a]	−0.82
TP-B and protein %	0.75	0.86	[a]	[a]	[a]	0.91
IgG and fat %	0.58	−0.08	[a]	[a]	[a]	0.40
IgG and lactose %	−0.80	−0.69	[a]	[a]	[a]	−0.88
IgG and protein %	0.84	0.87	[a]	[a]	[a]	0.92
Fat % and lactose %	−0.36	−0.16	[a]	[a]	[a]	−0.46
Fat % and protein %	0.60	−0.09	[a]	[a]	[a]	0.52
Lactose % and protein %	−0.87	−0.81	[a]	[a]	[a]	−0.92

Colostrum is taken d0 or d1 postpartum, milk is bulk tank milk from d5 or later. Total solids measured by Brix refractometer; IgG = immunoglobulin G; TP-B = total protein measured by Bradford assay.
[a]Data not available.

and Kummer, 1997), which may confound direct comparisons of the correlations. Notably the correlations between Brix refractometry and the components of individual cow colostrum from days 2 and 3 and in the bulk tank milk were low or negligible (Table 2). These results are similar to those of Rayburn *et al.* (2019), who found that refractometry was not useful for assessing IgG by the fourth or fifth milking. Therefore, using Brix refractometer or protein levels to estimate IgG in colostrum after 2 d postpartum is not advised.

IgG represents approximately 75% of the protein in colostrum (McGrath *et al.*, 2016; Godden *et al.*, 2019). However, other proteins are also important for calf health, including beta-lactoglobulin, alpha-lactalbumin, lactoferrin and other immunoglobulins (namely, IgA and IgM) (McGrath *et al.*, 2016). Although laboratory assays to measure colostrum components can be expensive and time consuming, they are highly accurate. Among these laboratory assays, the Bradford assay (TP-B) is a relatively easy and inexpensive method to measure total protein. The correlations between total protein, as measured by Bradford assay (TP-B) in the individual cow colostrum at day 0 and 1 postpartum, and IgG or protein % were high and positive ($r = 0.90$ and 0.91, respectively).

There was a strong and highly negative correlation between total protein (TP-B) and lactose ($r = -0.82$) (Table 2). The colostrum lactose concentration was also highly negatively correlated with protein % measured by FT-MIR ($r = -0.92$). This osmotic activity of lactose dilutes other colostrum components, specifically the protein concentration, resulting in an inverse relationship between the levels of lactose and protein in the colostrum. Consequently, the colostrum lactose concentration also had a high negative correlation with IgG concentration ($r = -0.88$), as has been observed by others (Dunn *et al.*, 2017).

Estimating colostrum components using Brix refractometer measurements

The very high correlations observed between each of the components of individual cow colostrum within 1 d postpartum and the pooled colostrum means that the concentration of one component can be potentially used to predict that of the others (Supplementary Figures S1–S4). The exception is fat percentage as it was not strongly correlated with the level of other colostrum components.

Simple linear model equations were used to estimate IgG, protein, and lactose concentrations based on the Brix refractometer measurements of individual cow colostrum day 0 and 1 postpartum and the pooled colostrum ($n = 29$). The very high correlations between the components were reflected in the coefficients of determination (R^2) (Supplementary Figure S1). Importantly, the formula for predicting IgG (IgG (g/l) = −103.1 + 10.9 × Brix %) had good predictive power ($R^2 = 0.80$, $P < 0.001$). Both r and R^2 in the present study were higher than reported other studies such as Bartier *et al.* (2015) (0.64 and 0.43, respectively) and Bielmann *et al.* (2010) (0.73 and 0.53, respectively). This may be a result of the method used to measure the IgG concentration in the present study as the ELISA is more sensitive and accurate than the RID assays used in the other studies.

Both lactose and protein levels in the individual cow colostrum 0–1 d postpartum and pooled colostrum could be estimated with reasonable accuracy from Brix % (Supplementary Figure S1). The lactose concentration could be estimated using the formula, Lactose (%) = 5.7–0.2 × Brix %, with a reasonable goodness of fit ($R^2 = 0.78$, $P < 0.001$). The total protein in colostrum could be estimated from the formula, TP-B (g/l) = −61.6 + 8.8 × Brix %, with a high coefficient of determination ($R^2 = 0.85$, $P < 0.001$).

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The formula predicting the protein % in colostrum from Brix refractometry measurements (Protein % = −2.5 + 0.7 × Brix %) had an even greater coefficient of determination ($R^2 = 0.96$, $P < 0.001$) than the TP-B formula. Therefore, Brix refractometry measurements are a good predictor for not only estimating IgG in colostrum, but also protein and lactose concentrations.

Previous studies suggest calves should receive at least 10% of body weight of 50 g/l of IgG from colostrum within a few hours of birth (McGuirk and Collins, 2004; Godden, 2008; Bartier *et al.*, 2015). As it is difficult to measure IgG on-farm, it has been suggested that calves should receive a large volume of colostrum, equal to 10 to 12% of body weight (3 to 4 litres of colostrum) at their first feed (Moran, 2002; McGuirk and Collins, 2004; Godden *et al.*, 2019). Other authors investigating the transfer of passive immunity to calves fed different colostrum volumes at different times after birth have suggested specific colostrum volumes and specific feeding times (Williams *et al.*, 2014). For example, Osaka *et al.* (2014) recommended that calves consume ⩾3 litres of colostrum with IgG concentration >40 mg/ml within 6 h after birth. However, it is difficult to follow these recommendations as there is a large variation in individual cow colostrum IgG concentrations (Marnila and Korhonen, 2011; Quigley *et al.*, 2013; Dunn *et al.*, 2017), and other variation is inevitable due to factors such as the time of collection, breed and cow management (Bartier *et al.*, 2015; Schneider *et al.*, 2020).

Establishing the colostrum quality using Brix refractometry will enable the volume of colostrum that should be fed to the neonate to avoid FPIT to be more accurately determined. For example, colostrum with a Brix of 20% would have approximately 115 g/l of IgG (Supplementary Table S1), so calves weighing ≤40 kg at birth should receive ~ 2 litres of colostrum of this quality within 4 h postpartum to avoid FPIT. If the Brix % is lower than the recommended threshold of 18 to 23 Brix%, then more colostrum should be provided at the first feed. Based on the prediction equation, a Brix measurement of 14% would be equivalent to 50 g/l of IgG, and the producer would need to feed calves weighing ≤ 40 kg at least 4 litres of the colostrum to ensure the calves received sufficient IgG (Supplementary Table S1).

Calf blood components

Blood samples were taken from the calves prior to the first feed (day 0), and then after receiving the colostrum and milk, on days 1 and 7. The blood samples were analysed for glucose, GGT, total protein and IgG. Before the first feed, all colostrum derived components were at low levels in the calf blood, in particular, GGT and IgG (11.7 IU/l and 0.3 g/l, respectively) (Table 3). These values confirmed that calves had not suckled from their dam prior to sampling (Braun *et al.*, 1982). The concentrations of all components increased after feeding (day 1), but declined by day 7 (Table 3). The exception was the IgG concentration, which was higher at day 7 than day 1.

Correlations between calf blood components

There were negligible or low correlations between the calf blood components in the samples taken before the calves received colostrum (day 0; Table 4). On day 1, the correlations between total protein measured by refractometry (TP-R) and GGT and between GGT and IgG were high ($r = 0.71$ and $r = 0.72$, respectively), and the correlation between TP-R and IgG was moderate ($r = 0.61$). The correlations between TP-R and IgG and between GGT and IgG remained constant up to day 7 ($r = 0.67$ and $r = 0.62$, respectively), but the correlations between other components were substantially lower at later time points. A higher correlation between TP-R and IgG was observed by combining all the calf data from the first week postpartum than for any of the individual days ($r = 0.81$).

The correlation between total protein measured by serum refractometer (TP-R) and IgG for calf blood samples within 7 d postpartum was similar to those reported in other studies, which ranged between from 0.75 to 0.93 (Deelen *et al.*, 2014; Hernandez *et al.*, 2016; Renaud *et al.*, 2018; Wilm *et al.*, 2018). High correlations have been observed between serum IgG and Brix refractometer measurements of serum as well ($r = 0.79$–0.93) (Deelen *et al.*, 2014; Hernandez *et al.*, 2016). These authors also reported a very high correlation between the values from these two different types of refractometers, Brix and digital serum protein ($r = 0.97$–1.0). The estimated FPIT thresholds are 8.1% to 8.5% for the Brix refractometer and 50 to 55 g/l for the serum protein refractometer (Godden *et al.*, 2019).

There was a high correlation between serum IgG and plasma GGT at day 1 ($r = 0.72$), but the correlation was lower by day 7 ($r = 0.62$) (Table 4). These correlations are similar to those reported in a study of calves less than 11 d postpartum ($r = 0.65$–0.68) (Parish *et al.*, 1997). However, much lower correlations

Table 3. Descriptive statistics of calf blood measurements

Days postpartum	Glucose (mmol/l)	GGT (U/l)	TP-R (g/l)	TP-B (g/l)	IgG (g/l)
Day 0 ($n = 35$)					
Mean ± SE	3.8 ± 0.2	11.7 ± 0.7	40.4 ± 0.6	52.5 ± 1.6	0.3 ± 0.1
Range	1.6–6.5	6.2–26.2	34.0–46.0	38.3–80.0	0–1.7
Day 1 ($n = 35$)					
Mean ± SE	6.9 ± 0.2	1305.9 ± 163.5	57.7 ± 1.4	76.6 ± 2.7	23.4 ± 2.1
Range	4.3–10.2	89.5–4034.5	38.0–74.0	50.6–107.1	7.2–58.1
Day 7 ($n = 35$)					
Mean ± SE	6.0 ± 0.1	205.7 ± 22.8	55.1 ± 1.1	63.7 ± 1.5	27.1 ± 2.4
Range	4.7–7.9	25.4–581.6	42.0–70.0	47.8–87.0	7.3–57.5

TP-R = total protein by refractometry; TP-B = total protein by Bradford assay; Day 0 = within 4 h postpartum and before receiving colostrum; Days 1 and 7 after receiving colostrum or milk twice per day.

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Table 4. Correlations between calf blood components at various times postpartum

Correlated parameters	Calf data			
	Day 0 ($n = 35$)	Day 1 ($n = 35$)	Day 7 ($n = 35$)	Days 0, 1, 7 ($n = 105$)
Glucose and TP-R	−0.16	0.07	0.09	0.57
Glucose and TP-B	−0.21	0.01	0.18	0.45
Glucose and IgG	0.05	0.02	−0.11	0.50
Glucose and GGT	−0.03	0.02	0.24	0.45
TP-R and TP-B	0.44	0.17	0.10	0.54
TP-R and IgG	0.17	0.61	0.67	0.81
TP-R and GGT	0.03	0.71	0.46	0.65
TP-B and IgG	0.46	0.50	0.05	0.51
TP-B and GGT	0.07	0.28	0.06	0.55
IgG and GGT	−0.02	0.72	0.62	0.52

TP-R = total protein by refractometry; TP-B = total protein by Bradford assay; Day 0 = within 4 h postpartum but before receiving colostrum; Days 1 and 7 after receiving colostrum or milk twice per day.

have been found in other studies of calves less than 8 d postpartum ($r = 0.398$ and $r = 0.495$) (Jezek *et al.*, 2010; Vandeputte *et al.*, 2011).

Estimating calf blood components using serum refractometry

As the correlations between the calf blood IgG and TP-R and between GGT and TP-R were high, regression models were developed to estimate the serum IgG and plasma GGT based on the serum protein refractometer measurements (Supplementary Figures S3 and S4). To predict calf blood components more accurately, all the data within 7 d postpartum were combined and a formula was generated to predict serum IgG concentration from the protein refractometer measurements, IgG (g/l) = −49.1 + 1.3 × TP-R (g/l), which had a coefficient of determination of $R^2 = 0.65$ ($P < 0.001$, Supplementary Figure S3). The correlation between serum IgG and the protein refractometer measurements was higher in the present study ($r = 0.81$) than that reported by Parish *et al.* (1997) ($r = 0.68$), most likely due to the age differences between the calves (7 d *vs.* 11 d of age, respectively). Hence, the regression model from Parish *et al.* (1997), IgG = −2,562 + total protein (g/dl) × 670, is likely to have an lower goodness of fit for predicting serum IgG than the prediction equation developed here.

Gamma-glutamyl transferase (GGT)

GGT is found in very low concentrations in the blood of newborn calves but is rapidly absorbed from the colostrum before the gut closes, and hence, is a good indicator of colostrum ingestion (Braun *et al.*, 1982; Weaver *et al.*, 2000). The level of GGT has been also proposed as an indicator of passive immunity transfer (Parish *et al.*, 1997; Wilson *et al.*, 1999; Aydogdu and Guzelbektes, 2018). Different GGT thresholds for FPIT have been suggested (Perino *et al.*, 1993; Parish *et al.*, 1997; Hogan *et al.*, 2015; Cuttance *et al.*, 2017*a*), but the threshold is yet to be firmly established (Cuttance *et al.*, 2019).

In order to estimate a FPIT threshold for GGT in the present study, a prediction equation for IgG concentration was generated based on GGT levels at day 1, GGT (U/l) = −3.9 + 55.9 × IgG (g/l) (Supplementary Figure S4). Assuming the FPIT threshold is 10 g/l IgG at day 1 postpartum, the FPIT threshold for GGT level is predicted from this equation to be approximately 555 IU/l. This level is similar to the experimental results from a study by Pekcan *et al.* (2013), who found that calves less than 2 d old with adequate passive immunity transfer had a mean of 578.25 ± 47.45 IU GGT/l. However, the prediction equation based on GGT level at day 1 postpartum only had a coefficient of determination of $R^2 = 0.52$ ($P < 0.001$, Supplementary Figure S4), and this estimate of FTP threshold for GGT is not expected to be highly accurate. In addition, the lower correlations and coefficients of determination indicate that plasma GGT activity was not prognostic of IgG concentration, particularly after day 1.

In conclusion, the results presented here validate both Brix and serum refractometry as good methods to estimate colostrum and calf blood components, respectively. Refractometry can be used to adjust the amount of colostrum fed to ensure calves receive sufficient IgG and nutrients from the colostrum or to predict IgG levels in calves and hence, the risk of FPIT. Nevertheless, other factors, such as the timing of the first feed, calf health and body weight, need to be considered to ensure sufficient transfer of passive immunity.

Supplementary material. The supplementary material for this article can be found at https://doi.org/10.1017/S0022029921000340

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